# Package 'FELLA'

November 27, 2024

Type Package

Title Interpretation and enrichment for metabolomics data

Version 1.27.0

Date 2019-10-19

Maintainer Sergio Picart-Armada <sergi.picart@upc.edu>

**Description** Enrichment of metabolomics data using KEGG entries. Given a set of affected compounds, FELLA suggests affected reactions, enzymes, modules and pathways using label propagation in a knowledge model network. The resulting subnetwork can be visualised and exported.

License GPL-3

LazyLoad yes

**Imports** methods, igraph, Matrix, KEGGREST, plyr, stats, graphics, utils

**Depends** R (>= 3.5.0)

Suggests shiny, DT, magrittr, visNetwork, knitr, BiocStyle, rmarkdown, testthat, biomaRt, org.Hs.eg.db, org.Mm.eg.db, AnnotationDbi, GOSemSim

VignetteBuilder knitr

RoxygenNote 6.1.1

Encoding UTF-8

**biocViews** Software, Metabolomics, GraphAndNetwork, KEGG, GO, Pathways, Network, NetworkEnrichment

Collate 'AllArguments.R' 'AllClasses.R' 'AllMethods.R' 'generateResultsTable.R' 'generateEnzymesTable.R' 'generateResultsGraph.R' 'exportResults.R' 'addGOToGraph.R' 'buildGraphFromKEGGREST.R' 'buildDataFromGraph.R' 'defineCompounds.R' 'doc-data.R' 'doc-package.R' 'runHypergeom.R' 'runDiffusion.R' 'runPagerank.R' 'enrich.R' 'get-.R' 'is-.R' 'launchApp.R' 'list-.R' 'loadKEGGdata.R' 'plotBipartite.R' 'plotGraph.R' 'plotLegend.R'

## Contents

git\_url https://git.bioconductor.org/packages/FELLA

git\_branch devel

git\_last\_commit d1ba32c

git\_last\_commit\_date 2024-10-29

**Repository** Bioconductor 3.21

Date/Publication 2024-11-26

Author Sergio Picart-Armada [aut, cre], Francesc Fernandez-Albert [aut], Alexandre Perera-Lluna [aut]

# Contents

params		3
checkArguments	 	 5
D.diffusion-class	 •	 6
D.hypergeom-class	 •	 7
D.keggdata-class	 •	 7
D.pagerank-class	 •	 7
lata-funs		8
enrich-funs	 •	 11
export-funs	 •	 16
FELLA	 •	 21
FELLA.DATA-class	 •	 22
FELLA.sample		23
FELLA.USER-class	 •	 24
getBackground	 •	 25
getCom		26
getExcluded	 	 27
getGraph	 	 27
getInfo	 	 28
getInput	 	 29
getMatrix	 	 29
getName	 	 30
getPscores	 	 31
getPvaluesSize	 	 31
getStatus	 	 32
getSums	 •	 33
getValid	 	 33
nfere.con2ec	 •	 34
nput.sample		35
s.FELLA.DATA	 •	 36
s.FELLA.USER	 •	 36
argestcc	 •	 37
aunchApp	 •	 38
istApprox	 •	 38
istCategories	 •	 39

## .params

listInternalDatabases	3	9
listMethods	4	0
mytriangle	4	0
plotBipartite	4	1
plotLegend	4	2
sanitise	4	.3
U.diffusion-class	4	.3
U.hypergeom-class	4	4
U.pagerank-class	4	4
U.userinput-class	4	5
	4	6

# Index

.params

Dummy function with function arguments

# Description

This function eases parameter inheritance to centralise the documentation

# Usage

.params()

## Arguments

databaseDir	Path for the KEGG RData files	
internalDir	Logical, is the directory located in the package directory?	
object	FELLA.USER object	
data	FELLA.DATA object	
type	Character vector, containing entries in "hypergeom", "diffusion" or "pagerank"	
level	Desired level, can be coded as a number or a character: 1 or "pathway"; 2 or "module"; 3 or "enzyme"; 4 or "reaction"; 5 or "compound".	
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"	
methods	Character vector, containing some of: "hypergeom", "diffusion", "pagerank"	
approx	Character: "simulation" for Monte Carlo, "normality", "gamma" or "t" for para- metric approaches	
loadMatrix	Character vector to choose if heavy matrices should be loaded. Can contain: "diffusion", "pagerank"	
threshold	Numeric value between 0 and 1. p.score threshold applied when filtering KEGG nodes. Lower thresholds are more stringent.	
thresholdConnectedComponent		
	Numeric value between 0 and 1. Connected components that are below the threshold are kept, while the ones exceeding it (because they are too small) are discarded.	

plimit	Pathway limit, must be a numeric value between 1 and 50. Limits the amount of pathways in method = "hypergeom"	
nlimit	Node limit, must be a numeric value between 1 and 1000. Limits the order of the solution sub-graph when in method = "diffusion" and method = "pagerank"	
niter	Number of iterations (permutations) for Monte Carlo ("simulation"), must be a numeric value between 1e2 and 1e5	
layout	Logical, should the plot be returned as a layout?	
graph	An <b>igraph</b> object, typically a small one, coming from an enrichment through "diffusion" or "pagerank".	
GOterm	Character, GO entry to draw semantic similarity in the solution graph. If NULL, the GO labels will be appended without similarities.	
GONamesAsLabels		
	Logical, should GO names be displayed as labels instead of GO identifiers?	
LabelLengthAtPl		
	Numeric value between 10 and 50. Maximum length that a label can reach when plotting the graph. The remaining characters will be truncated using ""	
godata.options	List, options for the database creator godata	
mart.options	List, options for the biomaRt function getBM. Importantly, this defines the or- ganism, see listDatasets for possibilities. If calling generateEnzymesTable, the user can set mart.options = NULL to avoid adding GO labels to enzymes.	
p.adjust	Character passed to the p.adjust method	
dampingFactor	Numeric value between 0 and 1 (none inclusive), damping factor d for PageRank (page.rank)	
t.df	Numeric value; number of degrees of freedom of the t distribution if the approx- imation approx = "t" is used	
compounds	Character vector containing the KEGG IDs of the compounds considered as affected	
compoundsBackground		
	Character vector containing the KEGG IDs of the compounds that belong to the background. Can be NULL for the default background (all compounds)	
NamesAsLabels	Logical, should KEGG names be displayed as labels instead of KEGG identifiers?	
capPscores	Numeric value, minimum p-score admitted for the readable formatting. Smaller p-scores will be displayed as < capPscores	

## Value

NULL

checkArguments

#### Description

This function checks if the arguments are of the desired type, length and range. If it fails, it gives an error explaining why the argument is invalid.

## Usage

```
checkArguments(databaseDir = "myDatabase", internalDir = TRUE,
  method = "diffusion", methods = "diffusion", approx = "normality",
  loadMatrix = NULL, threshold = 0.05, plimit = 15, nlimit = 250,
  niter = 1000, t.df = 10, dampingFactor = 0.85, layout = FALSE,
  thresholdConnectedComponent = 0.05, GOterm = NULL,
  GONamesAsLabels = TRUE, LabelLengthAtPlot = 22,
  object = new("FELLA.USER"), data = new("FELLA.DATA"), ...)
```

#### Arguments

databaseDir	Path for the KEGG RData files
internalDir	Logical, is the directory located in the package directory?
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"
methods	Character vector, containing some of: "hypergeom", "diffusion", "pagerank"
approx	Character: "simulation" for Monte Carlo, "normality", "gamma" or "t" for para- metric approaches
loadMatrix	Character vector to choose if heavy matrices should be loaded. Can contain: "diffusion", "pagerank"
threshold	Numeric value between 0 and 1. p.score threshold applied when filtering KEGG nodes. Lower thresholds are more stringent.
plimit	Pathway limit, must be a numeric value between 1 and 50. Limits the amount of pathways in method = "hypergeom"
nlimit	Node limit, must be a numeric value between 1 and 1000. Limits the order of the solution sub-graph when in method = "diffusion" and method = "pagerank"
niter	Number of iterations (permutations) for Monte Carlo ("simulation"), must be a numeric value between 1e2 and 1e5
t.df	Numeric value; number of degrees of freedom of the t distribution if the approx- imation approx = "t" is used
dampingFactor	Numeric value between 0 and 1 (none inclusive), damping factor d for PageRank (page.rank)
layout	Logical, should the plot be returned as a layout?

thresholdConnec	Numeric value between 0 and 1. Connected components that are below the threshold are kept, while the ones exceeding it (because they are too small) are discarded.	
GOterm	Character, GO entry to draw semantic similarity in the solution graph. If NULL, the GO labels will be appended without similarities.	
GONamesAsLabels	5	
	Logical, should GO names be displayed as labels instead of GO identifiers?	
LabelLengthAtPlot		
	Numeric value between 10 and 50. Maximum length that a label can reach when plotting the graph. The remaining characters will be truncated using ""	
object	FELLA.USER object	
data	FELLA.DATA object	
	ignored arguments	

#### Value

A list with values. Currently only a logical value named valid if the process runs smoothly. If the checking fails, it also returns an object called ans, which depends on the situation (can be the original object, NULL, et cetera).

#### Examples

```
## This function is internal
arg1 <- FELLA:::checkArguments(method = "hello")
arg1$valid
arg2 <- FELLA:::checkArguments(method = "diffusion")
arg2$valid</pre>
```

D.diffusion-class An internal S4 class for the diffusion data

## Description

An internal S4 class for the diffusion data

#### Slots

matrix Numeric (dense) matrix [optional] rowSums Numeric named vector with rowSums internal data squaredRowSums Numeric named vector with squaredRowSums internal data D.hypergeom-class An internal S4 class for the binary matrix (hypergeometric test)

#### Description

An internal S4 class for the binary matrix (hypergeometric test)

## Slots

matrix Binary sparse matrix

D.keggdata-class An internal S4 class to represent the KEGG graph and related files

#### Description

An internal S4 class to represent the KEGG graph and related files

## Slots

graph KEGG graph id2name Mapping list: KEGG ID to KEGG name (can contain multiple hits) pvalues.size Numeric matrix for the evaluation of CC through their size id List with character vectors for KEGG categories status Character that specifies the current status of this S4 class

D.pagerank-class An internal S4 class for the PageRank data

#### Description

An internal S4 class for the PageRank data

#### Slots

matrix Numeric (dense) matrix [optional] rowSums Numeric named vector with rowSums internal data squaredRowSums Numeric named vector with squaredRowSums internal data data-funs

#### Description

Function buildGraphFromKEGGREST makes use of the KEGG REST API (requires internet connection) to build and return the curated KEGG graph.

Function buildDataFromGraph takes as input the KEGG graph generated by buildGraphFromKEGGREST and writes the KEGG knowledge model in the desired permanent directory.

Function loadKEGGdata loads the internal files containing the KEGG knowledge model into a FELLA.DATA object.

In general, generateGraphFromKEGGREST and generateDataFromGraph are one-time executions for a given organism and knowledge model, in this precise order. On the other hand, the user needs to run loadKEGGdata in every new R session to load such model into a FELLA.DATA object.

#### Usage

```
buildGraphFromKEGGREST(organism = "hsa", filter.path = NULL)
```

```
buildDataFromGraph(keggdata.graph = NULL, databaseDir = NULL,
    internalDir = TRUE, matrices = c("hypergeom", "diffusion",
    "pagerank"), normality = c("diffusion", "pagerank"),
    dampingFactor = 0.85, niter = 100)
```

#### Arguments

organism	Character, KEGG code for the organism of interest
filter.path	Character vector, pathways to filter. This is a pattern matched using regexp. E.g: "01100" to filter the overview metabolic pathway in any species
keggdata.graph	An <b>igraph</b> object generated by the function buildGraphFromKEGGREST
databaseDir	Character containing the directory to save KEGG files. It is a relative directory inside the library location if internalDir = TRUE. If left to NULL, an automatic name containing the date, organism and the KEGG release is generated.
internalDir	Logical, should the directory be internal in the package directory?
matrices	A character vector, containing any of these: "hypergeom", "diffusion", "pagerank"
normality	A character vector, containing any of these: "diffusion", "pagerank"
dampingFactor	Numeric value between 0 and 1 (none inclusive), damping factor d for PageRank (page.rank)
niter	Numeric value, number of iterations to estimate the p-values for the CC size. Between 10 and 1e3.
loadMatrix	Character vector to choose if heavy matrices should be loaded. Can contain: "diffusion", "pagerank"

#### data-funs

#### Details

In function buildGraphFromKEGGREST, The user specifies (i) an organism, and (ii) patterns matching pathways that should not be included as nodes. A graph object, as described in [Picart-Armada, 2017], is built from the comprehensive KEGG database [Kanehisa, 2017]. As described in the main vignette, accessible through browseVignettes("FELLA"), this graph has five levels that represent categories of KEGG nodes. From top to bottom: pathways, modules, enzymes, reactions and compounds. This knowledge representation is resemblant to the one formerly used by MetScape [Karnovsky, 2011], in which enzymes connect to genes instead of modules and pathways. The necessary KEGG annotations are retrieved through KEGGREST R package [Tenenbaum, 2013]. Connections between pathways/modules and enzymes are inferred through organism-specific genes, i.e. an edge is added if a gene connects both entries. However, in order to enrich metabolomics data, the user has to pass the graph object to buildDataFromGraph to obtain the FELLA.USER object. All the networks are handled with the igraph R package [Csardi, 2006].

Using buildDataFromGraph is the second step to use the FELLA package. The knoledge graph is used to compute other internal variables that are required to run any enrichment. The main point behind the enrichment is to provide a small part of the knowledge graph relevant to the supplied metabolites. This is accomplished through diffusion processes and random walks, followed by a statistical normalisation, as described in [Picart-Armada, 2017]. When building the internal files, the user can choose whether to store (i) matrices for each provided method, and (ii) vectors derived from such matrices to use the parametric approaches. These are optional but enable (i) faster permutations and custom metabolite backgrounds, and (ii) parametric approaches. WARNING: diffusion and PageRank matrices in (i) can allocate up to 250MB each. On the other hand, the niter parameter controls the amount of trials to approximate the distribution of the connected component size under uniform node sampling. For further info, see the option thresholdConnectedComponent in the details from ?generateResultsGraph. Regarding the destination, the user can specify the name of the directory. Otherwise a name containing the creation date, the organism and the KEGG release will be used. The database can be stored within the library path or in a custom location.

Function loadKEGGdata returns a FELLA.DATA object from any of the databases generated by FELLA.DATA. This object is the starting point of any enrichment using FELLA. In case the user built the matrices for "diffusion" and "pagerank", he or she can choose to load them. Further detail on the methods can be found in [Picart-Armada, 2017]. The matrices allow a faster computation and the definition of a custom background, but use up to 250MB of memory each.

#### Value

buildGraphFromKEGGREST returns the curated KEGG graph (class igraph)

buildDataFromGraph returns invisible(TRUE) if successful. As a side effect, the directory outdir is created, containing the internal data.

loadKEGGdata returns the FELLA. DATA object that contains the KEGG knowledge representation.

#### References

Kanehisa, M., Furumichi, M., Tanabe, M., Sato, Y., & Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic acids research, 45(D1), D353-D361.

Karnovsky, A., Weymouth, T., Hull, T., Tarcea, V. G., Scardoni, G., Laudanna, C., ... & Athey, B. (2011). Metscape 2 bioinformatics tool for the analysis and visualization of metabolomics and gene expression data. Bioinformatics, 28(3), 373-380.

Tenenbaum, D. (2013). KEGGREST: Client-side REST access to KEGG. R package version, 1(1). Chang, W., Cheng, J., Allaire, JJ., Xie, Y., & McPherson, J. (2017). shiny: Web Application Framework for R. R package version 1.0.5. https://CRAN.R-project.org/package=shiny

Picart-Armada, S., Fernandez-Albert, F., Vinaixa, M., Rodriguez, M. A., Aivio, S., Stracker, T. H., Yanes, O., & Perera-Lluna, A. (2017). Null diffusion-based enrichment for metabolomics data. PLOS ONE, 12(12), e0189012.

#### See Also

class FELLA. DATA

#### Examples

```
## Toy example
## In this case, the graph is not built from current KEGG.
## It is loaded from sample data in FELLA
data("FELLA.sample")
## Graph to build the database (this example is a bit hacky)
g.sample <- FELLA:::getGraph(FELLA.sample)</pre>
dir.tmp <- paste0(tempdir(), "/", paste(sample(letters), collapse = ""))</pre>
## Build internal files in a temporary directory
buildDataFromGraph(
keggdata.graph = g.sample,
databaseDir = dir.tmp,
internalDir = FALSE,
matrices = NULL,
normality = NULL,
dampingFactor = 0.85,
niter = 10)
## Load database
myFELLA.DATA <- loadKEGGdata(</pre>
dir.tmp,
internalDir = FALSE)
myFELLA.DATA
## Not run:
## Full example
## First step: graph for Mus musculus discarding the mmu01100 pathway
## (an analog example can be built from human using organism = "hsa")
g.mmu <- buildGraphFromKEGGREST(</pre>
organism = "mmu",
filter.path = "mmu01100")
summary(g.mmu)
cat(comment(g.mmu))
## Second step: build internal files for this graph
## (consumes some time and memory, especially if we compute
"diffusion" and "pagerank" matrices)
buildDataFromGraph(
```

10

#### enrich-funs

```
keggdata.graph = g.mmu,
databaseDir = "example_db_mmu",
internalDir = TRUE,
matrices = c("hypergeom", "diffusion", "pagerank"),
normality = c("diffusion", "pagerank"),
dampingFactor = 0.85,
niter = 1e3)
## Third step: load the internal files into a FELLA.DATA object
FELLA.DATA.mmu <- loadKEGGdata(
"example_db_mmu",
internalDir = TRUE,
loadMatrix = c("diffusion", "pagerank"))
FELLA.DATA.mmu
```

```
## End(Not run)
```

enrich-funs

Functions to map and enrich a list of metabolites

#### Description

Function defineCompounds creates a FELLA. USER object from a list of compounds and a FELLA. DATA object.

Functions runHypergeom, runDiffusion and runPagerank perform an enrichment on a FELLA. USER with the mapped input metabolites (through defineCompounds) and a FELLA.DATA object. They are based on the hypergeometric test, the heat diffusion model and the PageRank algorithm, respectively.

Function enrich is a wrapper with the following order: loadKEGGdata (optional), defineCompounds and one or more in runHypergeom, runDiffusion and runPagerank

#### Usage

#### Arguments

compounds	Character vector containing the KEGG IDs of the compounds considered as affected
compoundsBackg	round
	Character vector containing the KEGG IDs of the compounds that belong to the background. Can be NULL for the default background (all compounds)
data	FELLA.DATA object
object	FELLA.USER object
p.adjust	Character passed to the p.adjust method
approx	Character: "simulation" for Monte Carlo, "normality", "gamma" or "t" for para- metric approaches
t.df	Numeric value; number of degrees of freedom of the t distribution if the approx- imation approx = "t" is used
niter	Number of iterations (permutations) for Monte Carlo ("simulation"), must be a numeric value between 1e2 and 1e5
dampingFactor	Numeric value between 0 and 1 (none inclusive), damping factor d for PageRank (page.rank)
methods	Character vector, containing some of: "hypergeom", "diffusion", "pagerank"
loadMatrix	Character vector to choose if heavy matrices should be loaded. Can contain: "diffusion", "pagerank"
databaseDir	Character, path to load the FELLA. DATA object if it is not already passed through the argument data
internalDir	Logical, is the directory located in the package directory?
	Further arguments for the enrichment $function(s)\ \mbox{runDiffusion},\ \mbox{runPagerank}$

## Details

Function defineCompounds maps the specificied list of KEGG compounds [Kanehisa, 2017], usually from an experimental metabolomics study, to the graph contained in the FELLA.DATA object. Importantly, the names must be KEGG ids, so other formats (common names, HMDB ids, etc) must be mapped to KEGG first. For example, through the "Compound ID Conversion" tool in MetaboAnalyst [Xia, 2015]. The user can also define a personalised background as a list of KEGG compound ids, which should be more extensive than the list of input metabolites. Once the compounds are mapped, the enrichment can be performed through runHypergeom, runDiffusion and runPagerank.

Function runHypergeom performs an over representation analysis through the hypergeometric test [Fisher, 1935] on a FELLA.USER object with mapped metabolites and a FELLA.DATA object. If a custom background was specified, it will be used. This approach is included for completeness and it is not the main purpose behind the FELLA package. Importantly, runHypergeom is not a hypergeometric test using the original KEGG pathways. Instead, a compound "belongs" to a "pathway" if it can reach the original pathway in the upwards-directed KEGG graph. This is a way to evaluate enrichment including indirect connections to a pathway, e.g. through an enzymatic family. New "pathways" are expected to be larger than the original pathways in this analysis and therefore the results can differ from the standard over representation.

Function runDiffusion performs the diffusion-based enrichment on a FELLA.USER object with mapped metabolites and a FELLA.DATA object [Picart-Armada, 2017]. If a custom background was specified, it will be used. The idea behind the heat diffusion is the usage of the finite difference formulation of the heat equation to propagate labels from the metabolites to the rest of the graph.

Following the notation in [Picart-Armada, 2017], the temperatures (diffusion scores) are computed as:

$$T = -KI^{-1} \cdot G$$

G is an indicator vector of the input metabolites (1 if input metabolite, 0 otherwise). KI is the matrix -KI = L + B, being L the unnormalised graph Laplacian and B the diagonal matrix with B[i,i] = 1 if node i is a pathway and B[i,i] = 0 otherwise.

Equivalently, with the notation in the HotNet approach [Vandin, 2011], the stationary temperature is named fs:

$$f^s = L_{\gamma}^{-1} \cdot b^s$$

bs is the indicator vector G from above. Lgamma, on the other hand, is found as Lgamma =  $L + gamma \times I$ , where L is the unnormalised graph Laplacian, gamma is the first order leaking rate and I is the identity matrix. In our formulation, only the pathway nodes are allowed to leak, therefore I is switched to B. The parameter gamma is set to gamma = 1.

The input metabolites are forced to stay warm, propagating flow to all the nodes in the network. However, only pathway nodes are allowed to evacuate this flow, so that its directionality is bottomup. Further details on the setup of the diffusion process can be found in the supplementary file S2 from [Picart-Armada, 2017].

Finally, the warmest nodes in the graph are reported as the relevant sub-network. This will probably include some input metabolites and also reactions, enzymes, modules and pathways. Other metabolites can be suggested as well.

Function runPagerank performs the random walk based enrichment on a FELLA.USER object with mapped metabolites and a FELLA.DATA object. If a custom background was specified, it will be used. PageRank was originally conceived as a scoring system for websites [Page, 1999]. Intuitively, PageRank favours nodes that (1) have a large amount of nodes pointing at them, and (2) whose pointing nodes also have high scores. Classical PageRank is formulated in terms of a random walker - the PageRank of a given node is the stationary probability of the walker visiting it.

The walker chooses, in each step, whether to continue the random walk with probability dampingFactor or to restart it with probability 1 - dampingFactor. In the original publication, dampingFactor = 0.85, which is the value used in FELLA by default. If he or she continues, an edge is picked from the outgoing edges in the current node with a probability proportional to its weight. If he or she restarts it, a node is uniformly picked from the whole graph. The "personalised PageRank" variant allows a user-defined distribution as the source of new random walks. The R package igraph contains such variant in its page.rank function [Csardi, 2006].

As described in the supplement S3 from [Picart-Armada, 2017], the PageRank PR can be computed as a column vector by imposing a stationary state in the probability. With a damping factor d and the user-defined distribution p as a column vector:

$$\mathbf{PR} = d \cdot M \cdot \mathbf{PR} + (1 - d) \cdot p$$

M is the matrix whose element M[i,j] is the probability of transitioning from j to i. If node j has outgoing edges, their probability is proportional to their weight - all weights must be positive. If node j has no outgoing edges, the probability is uniform over all the nodes, i.e. M[i,j] = 1/nrow(M) for every i. Note that all the columns from M sum up exactly 1. This leads to an expression to compute PageRank:

$$\mathbf{PR} = (1-d)p \cdot (I-dM)^{-1}$$

The idea behind the method "pagerank" is closely related to "diffusion". Relevant metabolites are the sources of new random walks and nodes are scored through their PageRank. Specifically, p is set to a uniform probability on the input metabolites. More details on the setup can be found in the supplementary file S3 from [Picart-Armada, 2017].

There is an important detail for "diffusion" and "pagerank": the scores are statistically normalised. Omitting this normalisation leads to a systematic bias, especially in pathway nodes, as described in [Picart-Armada, 2017].

Therefore, in both cases, scores undergo a normalisation through permutation analysis. The score of a node i is compared to its null distribution under input permutation, leading to their p-scores. As described in [Picart-Armada, 2017], two alternatives are offered: a parametric and deterministic approach and a non-parametric, stochastic one.

Stochastic Monte Carlo trials ("simulation") imply randomly permuting the input niter times and counting, for each node i, how many trials led to an equally or more extreme value than the original score. An empirical p-value is returned [North, 2002].

On the other hand, the parametric scores (approx = "normality") give a z-score for such permutation analysis. The expected value and variance of such null distributions are known quantities, see supplementary file S4 from [Picart-Armada, 2017]. To work in the same range [0,1], z-scores are transformed using the routine pnorm. The user can also choose the Student's t using approx = "t" and choosing a number of degrees of freedom through t.df. This uses the function pt instead. Alternatively, a gamma distribution can be used by setting approx = "gamma". The theoretical mean (E) and variance (V) are used to define the shape (E^2/V) and scale (V/E) of the gamma distribution, and pgamma to map to [0,1].

Any sub-network prioritised by "diffusion" and "pagerank" is selected by applying a threshold on the p-scores.

Finally, the function enrich is a wrapper to perform the enrichment analysis. If no FELLA.DATA object is supplied, it loads it, maps the affected compounds and performs the desired enrichment(s) with a single call. Returned is a list with the loaded FELLA.DATA object and the results in a FELLA.USER object. Conversely, the user can supply the FELLA.DATA object and the wrapper will map the metabolites and run the desired enrichment method(s). In this case, only the FELLA.USER will be returned.

#### Value

defineCompounds returns the FELLA.USER object with the mapped metabolites, ready to be enriched.

runHypergeom returns a FELLA. USER object updated with the hypergeometric test results

runDiffusion returns a FELLA. USER object updated with the diffusion enrichment results

runPagerank returns a FELLA. USER object updated with the PageRank enrichment results

#### enrich-funs

enrich returns a FELLA. USER object updated with the desired enrichment results if the FELLA. DATA was supplied. Otherwise, a list with the freshly loaded FELLA.DATA object and the corresponding enrichment in the FELLA.USER object.

#### References

Kanehisa, M., Furumichi, M., Tanabe, M., Sato, Y., & Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic acids research, 45(D1), D353-D361.

Xia, J., Sinelnikov, I. V., Han, B., & Wishart, D. S. (2015). MetaboAnalyst 3.0 - making metabolomics more meaningful. Nucleic acids research, 43(W1), W251-W257.

Fisher, R. A. (1935). The logic of inductive inference. Journal of the Royal Statistical Society, 98(1), 39-82.

Picart-Armada, S., Fernandez-Albert, F., Vinaixa, M., Rodriguez, M. A., Aivio, S., Stracker, T. H., Yanes, O., & Perera-Lluna, A. (2017). Null diffusion-based enrichment for metabolomics data. PLOS ONE, 12(12), e0189012.

Vandin, F., Upfal, E., & Raphael, B. J. (2011). Algorithms for detecting significantly mutated pathways in cancer. Journal of Computational Biology, 18(3), 507-522.

Page, L., Brin, S., Motwani, R., & Winograd, T. (1999). The PageRank citation ranking: Bringing order to the web. Stanford InfoLab.

Csardi, G., & Nepusz, T. (2006). The igraph software package for complex network research. InterJournal, Complex Systems, 1695(5), 1-9.

North, B. V., Curtis, D., & Sham, P. C. (2002). A note on the calculation of empirical P values from Monte Carlo procedures. American journal of human genetics, 71(2), 439.

#### Examples

```
## Load the internal database.
## This one is a toy example!
## Do not use as a regular database
data(FELLA.sample)
## Load a list of compounds to enrich
data(input.sample)
```

```
## First, map the compounds
obj <- defineCompounds(
  compounds = c(input.sample, "I_dont_map", "me_neither"),
  data = FELLA.sample)
obj
## See the mapped and unmapped compounds
getInput(obj)
getExcluded(obj)
## Compounds are already mapped
## We can enrich using any method now</pre>
```

```
## If no compounds are mapped an error is thrown. Example:
## Not run:
```

```
data(FELLA.sample)
obj <- defineCompounds(</pre>
compounds = c("C00049", "C00050"),
data = FELLA.sample)
## End(Not run)
## Enrich using hypergeometric test
obj <- runHypergeom(</pre>
object = obj,
data = FELLA.sample)
obj
## Enrich using diffusion
## Note how the results are added;
## the hypergeometric results are not overwritten
obj <- runDiffusion(</pre>
object = obj,
approx = "normality",
data = FELLA.sample)
obj
## Enrich using PageRank
## Again, this does not overwrite other methods
obj <- runPagerank(</pre>
object = obj,
approx = "simulation",
data = FELLA.sample)
obj
## Example using the "enrich" wrapper
## Only diffusion
obj.wrap <- enrich(</pre>
compounds = input.sample,
method = "diffusion",
data = FELLA.sample)
obj.wrap
## All the methods
obj.wrap <- enrich(</pre>
compounds = input.sample,
methods = FELLA::listMethods(),
data = FELLA.sample)
obj.wrap
```

export-funs

Generate and manipulate tables and sub-networks from an enrichment

16

#### export-funs

#### Description

In general, generateResultsTable, generateEnzymesTable and generateResultsGraph provide the results of an enrichment in several formats.

Function generateResultsTable returns a table that contains the best hits from a FELLA.USER object with a successful enrichment analysis. Similarly, generateEnzymesTable returns a data frame with the best scoring enzyme families and their annotated genes.

Function generateResultsGraph gives a sub-network, plottable through plotGraph, with the nodes with the lowest p.score from an enrichment analysis. Function addGOToGraph can be applied to such sub-networks to overlay GO labels and similarity to a user-defined GO term.

Function exportResults is a wrapper around generateResultsTable, generateEnzymesTable and generateResultsGraph to write the results to files.

#### Usage

```
generateResultsTable(method = "diffusion", threshold = 0.05,
   plimit = 15, nlimit = 250, LabelLengthAtPlot = 45,
    capPscores = 1e-06, object = NULL, data = NULL, ...)
generateEnzymesTable(method = "diffusion", threshold = 0.05,
   nlimit = 250, LabelLengthAtPlot = 45, capPscores = 1e-06,
   mart.options = list(biomart = "ensembl", dataset =
    "hsapiens_gene_ensembl"), object = NULL, data = NULL, ...)
generateResultsGraph(method = "diffusion", threshold = 0.05,
   plimit = 15, nlimit = 250, thresholdConnectedComponent = 0.05,
   LabelLengthAtPlot = 22, object = NULL, data = NULL, ...)
exportResults(format = "csv", file = "myOutput",
   method = "diffusion", object = NULL, data = NULL, ...)
addGOToGraph(graph = NULL, GOterm = NULL, godata.options = list(OrgDb
   = "org.Hs.eg.db", ont = "CC"), mart.options = list(biomart = "ensembl",
   dataset = "hsapiens_gene_ensembl"))
plotGraph(graph = NULL, layout = FALSE, graph.layout = NULL,
   plotLegend = TRUE, plot.fun = "plot.igraph", NamesAsLabels = TRUE,
    ...)
```

#### Arguments

method	one in "diffusion", "pagerank"
threshold	Numeric value between 0 and 1. p.score threshold applied when filtering KEGG nodes. Lower thresholds are more stringent.
plimit	Pathway limit, must be a numeric value between 1 and 50. Limits the amount of pathways in method = "hypergeom"
nlimit	Node limit, must be a numeric value between 1 and 1000. Limits the order of the solution sub-graph when in method = "diffusion" and method = "pagerank"

LabelLengthAtPl	ot
	Numeric value between 10 and 50. Maximum length that a label can reach when plotting the graph. The remaining characters will be truncated using ""
capPscores	Numeric value, minimum p-score admitted for the readable formatting. Smaller p-scores will be displayed as < capPscores
object	FELLA.USER object
data	FELLA.DATA object
	Optional arguments for the plotting function in plotGraph. Arguments passed to the exporting function in exportResults. Ignored otherwise.
mart.options	List, options for the biomaRt function getBM. Importantly, this defines the or- ganism, see listDatasets for possibilities. If calling generateEnzymesTable, the user can set mart.options = NULL to avoid adding GO labels to enzymes.
thresholdConnec	
	Numeric value between 0 and 1. Connected components that are below the threshold are kept, while the ones exceeding it (because they are too small) are discarded.
format	Character, one of: "csv" for regular results table, "enzyme" for table with en- zyme data, "igraph" for igraph format. Alternatively, any format supported by igraph, see write_graph
file	Character specifying the output file name
graph	An <b>igraph</b> object, typically a small one, coming from an enrichment through "diffusion" or "pagerank".
GOterm	Character, GO entry to draw semantic similarity in the solution graph. If NULL, the GO labels will be appended without similarities.
godata.options	List, options for the database creator godata
layout	Logical, should the plot be returned as a layout?
graph.layout	Two-column numeric matrix, if this argument is not null then it is used as graph layout
plotLegend	Logical, should the legend be plotted as well?
plot.fun	Character, can be either plot.igraph or tkplot
NamesAsLabels	Logical, should KEGG names be displayed as labels instead of KEGG identifiers?

#### Details

Functions generateResultsTable and generateEnzymesTable need a FELLA.DATA object and a FELLA.USER object with a successful enrichment. generateResultsTable provides the entries whose p-score is below the chosen threshold in a tabular format. generateEnzymesTable returns a table that contains (1) the enzymes that are below the user-defined p-score threshold, along with (2) the genes that belong to the enzymatic families in the organism defined in the database, and (3) GO labels of such enzymes, if mart.options is not NULL and points to the right database.

Function generateResultsGraph returns an **igraph** object with a relevant sub-network for manual examination. A FELLA.USER object with a successful enrichment analysis and the corresponding

#### export-funs

FELLA.DATA must be supplied. Graph nodes are prioritised by p.score and selected through the most stringent between (1) p.score threshold and (2) maximum number of nodes nlimit.

There is an additional filtering feature for tiny connected components, controllable through thresholdConnectedComponent (smaller is stricter). The user can choose to turn off this filter by setting thresholdConnectedComponent = 1. The idea is to discard connected components so small that are likely to arise from random selection of nodes. Let k be the order of the current sub-network. A connected component of order r will be kept only if the probability that a random subgraph from the whole KEGG knowledge model of order k contains a connected component of order at least r is smaller than thresholdConnectedComponent. Such probabilities are estimated during buildDataFromGraph; the amount of random trials can be controlled by its niter argument.

Function exportResults writes the enrichment results as the specified filetype. Options are: a csv table ("csv"), an enzyme csv table ("enzyme") an **igraph** object as an RData file, or any format supported by igraph's write\_graph.

Function addGOToGraph takes and returns a graph object with class **igraph** adding the following attributes: GO labels in V(graph)\$GO, and semantic similarities in V(graph)\$GO.simil if GOterm != NULL.

The GO database describes genes in terms of three ontologies: molecular function (MF), biological process (BP) and cellular component (CC) [Gene Ontology Consortium, 2015]. The user can be interested in finding which enzymatic families reported with a low p. score are closest to a particular GO term. To assess similarity between GO labels, FELLA uses the semantic similarity defined in [Yu, 2010] and their implementation in the **GOSemSim** R package. The user will obtain, for each enzymatic family, the closest GO term to his or her GO query and the semantic similarity between them. Exact matches have a similarity of 1. Function plotGraph detects the presence of the GO similarity option and plots its magnitude.

Function plotGraph plots a solution graph from the diffusion and pagerank analysis. For plotting hypergeom results, please use plot instead. Specific colors and shapes for each KEGG category are used: pathways are maroon, modules are violet, enzymes are orange, reactions are blue and compounds are green. If the graph contains the similarity to a GO term, enzymes will be displayed as triangles whose color depicts the strength of such measure (yellow: weak, purple: strong). At the moment, plotGraph allows plotting throug the static plot.igraph and the interactive tkplot.

#### Value

generateResultsTable returns a data.frame that contains the nodes below the p.score threshold from an enrichment analysis

generateEnzymesTable returns a data.frame that contains the enzymes below the p.score threshold, along with their genes and GO labels

generateResultsGraph returns an **igraph** object: a sub-network from the whole KEGG knowledge model under the specified thresholds (threshold and thresholdConnectedComponent)

exportResults returns invisible(), but as a side effect the specified file is created.

addGOToGraph returns an **igraph** object, which is the input graph with extra attributes: GO labels in V(graph)\$GO, and semantic similarities in V(graph)\$GO.simil if GOterm != NULL

plotGraph returns invisible() if layout = F and the plotting layout as a data.frame otherwise.

#### References

Gene Ontology Consortium. (2015). Gene ontology consortium: going forward. Nucleic acids research, 43(D1), D1049-D1056.

Yu, G., Li, F., Qin, Y., Bo, X., Wu, Y., & Wang, S. (2010). GOSemSim: an R package for measuring semantic similarity among GO terms and gene products. Bioinformatics, 26(7), 976-978.

## Examples

```
## First generate a toy enrichment
library(igraph)
data(FELLA.sample)
data(input.sample)
## Enrich input
obj <- enrich(
compounds = input.sample,
data = FELLA.sample)
```

#### 

```
## Results table
tab.res <- generateResultsTable(
method = "hypergeom",
threshold = 0.1,
object = obj,
data = FELLA.sample)
head(tab.res)</pre>
```

```
tab.res <- generateResultsTable(
method = "diffusion",
threshold = 0.1,
object = obj,
data = FELLA.sample)
head(tab.res)</pre>
```

#### 

```
## Use wrapper to write the table to a file
out.file <- tempfile()
exportResults(
format = "csv",
threshold = 0.1,
file = out.file,
object = obj,
data = FELLA.sample)
tab.wrap <- read.csv(out.file)
head(tab.wrap)</pre>
```

#### 

## Enzymes table
tab.ec <- generateEnzymesTable(
threshold = 0.1,
object = obj,
data = FELLA.sample,</pre>

## FELLA

mart.options = NULL) head(tab.ec) ## Generate graph g.res <- generateResultsGraph(</pre> method = "pagerank", threshold = 0.1, object = obj, data = FELLA.sample) g.res ## Plot graph (without GO terms) plotGraph(g.res) ## Add similarity to the GO CC term "mitochondrion" ## Not run: g.cc <- FELLA:::addGOToGraph(</pre> graph = g.res, GOterm = "GO:0005739") ## Plot graph (with GO terms) plotGraph(g.cc) ## Without the CC any(V(g.res)\$GO.simil >= 0) ## With the CC v.cc <- unlist(V(g.cc)\$GO.simil)</pre> sum(v.cc >= 0, na.rm = TRUE) ## Similarity values table(v.cc)

## End(Not run)

FELLA

The FELLA package

#### Description

FELLA is a metabolomics data enrichment tool that contextualises a list of metabolites using KEGG reactions, enzymes, modules and pathways [Picart-Armada, 2017].

## Details

FELLA can build knowledge models for the desired organism from the KEGG database [Kanehisa, 2017]. Once a model is ready, the input for the enrichment is introduced as a list of affected metabolites (as KEGG IDs). The output contains a comprehensive biological network layout that relates relevant pathways to the affected metabolites. Results are available in network and tabular format.

FELLA is equipped with a simple graphical interface for the lay user, deployed through launchApp.

FELLA relies mainly on the following packages: KEGGREST for the queries to the KEGG server [Tenenbaum, 2013], **igraph** for the network support [Csardi, 2006] and **shiny** for the graphical user interface [Chang, 2017].

#### References

Methodology:

Picart-Armada, S., Fernandez-Albert, F., Vinaixa, M., Rodriguez, M. A., Aivio, S., Stracker, T. H., Yanes, O., & Perera-Lluna, A. (2017). Null diffusion-based enrichment for metabolomics data. PLOS ONE, 12(12), e0189012.

Database:

Kanehisa, M., Furumichi, M., Tanabe, M., Sato, Y., & Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic acids research, 45(D1), D353-D361.

Main dependencies:

Tenenbaum, D. (2013). KEGGREST: Client-side REST access to KEGG. R package version, 1(1).

Csardi, G., & Nepusz, T. (2006). The igraph software package for complex network research. InterJournal, Complex Systems, 1695(5), 1-9.

Chang, W., Cheng, J., Allaire, JJ., Xie, Y., & McPherson, J. (2017). shiny: Web Application Framework for R. R package version 1.0.5. https://CRAN.R-project.org/package=shiny

#### Examples

```
## Walkthrough
browseVignettes("FELLA")
## I: create database
?buildGraphFromKEGGREST
## II: enrich data
?enrich
## III: export results
?exportResults
```

FELLA.DATA-class An S4 class to represent all the necessary KEGG data

#### Description

An S4 class to represent all the necessary KEGG data

"show" is an S4 method to show a FELLA.DATA object

#### Usage

## S4 method for signature 'FELLA.DATA'
show(object)

## FELLA.sample

#### Arguments

object A FELLA. DATA object

## Value

show returns invisible()

#### Slots

keggdata A D.keggdata S4 object hypergeom A D.hypergeom S4 object diffusion A D.diffusion S4 object pagerank A D.pagerank S4 object

FELLA.sample

#### FELLA.DATA sample data

#### Description

This FELLA.DATA object is a small KEGG graph object. Despite being a small database that only contains the two metabolic pathways hsa00010 - Glycolysis / Gluconeogenesis, and hsa00640 - Propanoate metabolism, it is useful to play around with FELLA's functions. It is also used for internal testing of this package.

#### Usage

data(FELLA.sample)

#### Format

An object of class FELLA. DATA of length 1.

#### Value

A FELLA. DATA object

#### Source

Generated from a mid-2017 KEGG release (http://www.genome.jp/kegg/)

## Examples

data(FELLA.sample)

FELLA.USER-class An S4 cla

## Description

Assigning the value of show to a variable will provide small data frames with the best scoring pathways (hypergeom) and the best nodes in the KEGG network (diffusion and pagerank)

## Usage

```
## S4 method for signature 'FELLA.USER'
show(object)
## S4 method for signature 'FELLA.USER,missing'
plot(x = new("FELLA.USER"),
    method = "hypergeom", threshold = 0.05, plimit = 15,
    nlimit = 250, layout = FALSE, thresholdConnectedComponent = 0.05,
    LabelLengthAtPlot = 22, data = NULL, ...)
```

## Arguments

A FELLA. USER object
A FELLA. USER object
Character, exactly one of: "hypergeom", "diffusion", "pagerank"
Numeric value between 0 and 1. p.score threshold applied when filtering KEGG nodes. Lower thresholds are more stringent.
Pathway limit, must be a numeric value between 1 and 50. Limits the amount of pathways in method = "hypergeom"
Node limit, must be a numeric value between 1 and 1000. Limits the order of the solution sub-graph when in method = "diffusion" and method = "pagerank"
Logical, should the plot be returned as a layout?
tedComponent
Numeric value between 0 and 1. Connected components that are below the threshold are kept, while the ones exceeding it (because they are too small) are discarded.
ot
Numeric value between 10 and 50. Maximum length that a label can reach when plotting the graph. The remaining characters will be truncated using ""
FELLA.DATA object
Additional arguments passed to plotting functions

## Value

show invisibly returns a list of data frames with the best hits for each applied method plot returns a layout if layout = T, otherwise invisible()

## getBackground

## Slots

userinput A U. userinput S4 object hypergeom A U. hypergeom S4 object diffusion A U. diffusion S4 object pagerank A U. pagerank S4 object

getBackground Get compounds in the defined background

## Description

Extractor function for the compounds defined as background

#### Usage

getBackground(object)

#### Arguments

object FELLA.USER object

#### Value

Vector of compounds in the background. If this vector is empty, all the compounds are used as background by default.

## Examples

```
data(FELLA.sample)
data(input.sample)
input <- head(input.sample, 12)</pre>
## If the background is default, we see an empty vector
## Note that the number of iterations is really small in the example
obj <- enrich(</pre>
compounds = input,
method = "diffusion",
approx = "simulation",
niter = 100,
data = FELLA.sample)
getBackground(obj)
## Otherwise we see the background compounds that mapped to the graph
obj <- enrich(</pre>
compounds = input,
compoundsBackground = input.sample,
method = "diffusion",
```

getCom

```
approx = "simulation",
niter = 100,
data = FELLA.sample)
getBackground(obj)
```

getCom

Get community

## Description

Extractor function for all the nodes from a level/community of KEGG graph

## Usage

```
getCom(data, level, format = "name")
```

## Arguments

data	FELLA.DATA object
level	Desired level, can be coded as a number or a character: 1 or "pathway"; 2 or "module"; 3 or "enzyme"; 4 or "reaction"; 5 or "compound".
format	Format of the output, "name" returns KEGG IDs whereas "id" returns vertices IDs

## Value

Vector of the names/ids of the desired KEGG graph community

## Examples

```
data(FELLA.sample)
## Pathways
getCom(FELLA.sample, 1, format = "name")
getCom(FELLA.sample, 1, format = "id")
## Modules
getCom(FELLA.sample, 2)
## Enzymes
head(getCom(FELLA.sample, 3))
## Reactions
head(getCom(FELLA.sample, 4))
## Compounds
head(getCom(FELLA.sample, 5))
```

26

getExcluded

#### Description

Extractor function for the compounds in the input that were not mapped to the KEGG graph

## Usage

```
getExcluded(object)
```

#### Arguments

object FELLA.USER object

## Value

Vector of the excluded compounds

#### Examples

data(FELLA.sample)
data(input.sample)

## No excluded compounds
obj <- defineCompounds(
compounds = input.sample,
data = FELLA.sample)
getExcluded(obj)</pre>

```
## One compound does not map
## The user gets a warning as well
obj <- defineCompounds(
  compounds = c(input.sample, "intruder"),
data = FELLA.sample)
getExcluded(obj)</pre>
```

getGraph

Get KEGG graph

## Description

Extractor function for the KEGG graph from the FELLA.DATA object

#### Usage

getGraph(data)

#### Arguments

data FELLA.DATA object

## Value

KEGG graph as an igraph object

# Examples

```
data(FELLA.sample)
g <- getGraph(FELLA.sample)
class(g)</pre>
```

```
getInfo
```

## Get KEGG version info

## Description

Extractor function for the info about the KEGG version used to build the FELLA.DATA object

## Usage

getInfo(data)

## Arguments

data FELLA.DATA object

#### Value

Character containing the KEGG release details

## Examples

```
data(FELLA.sample)
getInfo(FELLA.sample)
```

getInput

#### Description

Extractor function for the metabolites specified by the user in the input

#### Usage

```
getInput(object)
```

#### Arguments

object FELLA.USER object

## Value

Vector of metabolites in the input

#### Examples

```
data(FELLA.sample)
data(input.sample)
## No excluded compounds: the input is recovered as is
obj <- defineCompounds(
compounds = input.sample,
data = FELLA.sample)
i1 <- getInput(obj)
## One compound does not map: the input contains only the mapped entities
obj <- defineCompounds(
compounds = c(input.sample, "intruder"),
data = FELLA.sample)
i2 <- getInput(obj)
identical(sort(i1), sort(i2))</pre>
```

getMatrix

Get matrix for the desired methodology

#### Description

Extractor function for the matrices of hypergeometric, diffusion and PageRank methodologies

#### Usage

getMatrix(data, method)

getName

#### Arguments

data	FELLA.DATA object
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"

## Value

Matrix for the desired methodology (internal usage)

## Examples

```
## This function is internal
attach(environment(FELLA:::getMatrix))
data(FELLA.sample)
# When a matrix is loaded:
x <- getMatrix(FELLA.sample, "hypergeom")
dim(x)
# When it is not:
y <- getMatrix(FELLA.sample, "diffusion")
dim(y)
y</pre>
```

getName

Map KEGG identifiers to KEGG names

## Description

Map KEGG identifiers to KEGG names, multiple names for an ID are reported if annotated. The KEGG identifiers may have mixed levels.

## Usage

getName(data, id)

#### Arguments

data	FELLA.DATA object
id	KEGG IDs whose name is desired

## Value

List whose names are KEGG IDs and whose entries are the vectors of matches

#### Examples

```
data(FELLA.sample)
getName(FELLA.sample, c("C00002", "C00040"))
```

getPscores

## Description

Extractor function for the p-scores using the desired methodology

## Usage

```
getPscores(object, method)
```

## Arguments

object	FELLA.USER object
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"

## Value

Named vector of p-scores

## Examples

```
data(FELLA.sample)
data(input.sample)
obj <- enrich(
compounds = input.sample,
data = FELLA.sample)
p <- getPscores(obj, "diffusion")
sum(p < 0.1)</pre>
```

getPvaluesSize Get matrix for the p-value regarding CC size

## Description

Extractor function for the matrix containing p-value by CC size that compares to a random selection of nodes in the KEGG graph

## Usage

getPvaluesSize(data)

#### Arguments

data FELLA.DATA object

## getStatus

## Value

Matrix with p-values for CC size (internal usage)

#### Examples

```
## This function is internal
attach(environment(FELLA:::getPvaluesSize))
data(FELLA.sample)
M <- getPvaluesSize(FELLA.sample)
dim(M)
summary(as.vector(M))</pre>
```

getStatus

## Get the slot "status"

## Description

Extractor function for the slot "status" for the KEGG data

## Usage

getStatus(data)

#### Arguments

data FELLA.DATA object

## Value

Slot "status" (internal usage)

## Examples

## This function is internal

data(FELLA.sample)

## Is the object loade?
FELLA:::getStatus(FELLA.sample)
FELLA:::getStatus(new("FELLA.DATA"))

32

getSums

# Description

Extractor function for rowSums/squaredRowSums

## Usage

```
getSums(data, method, squared)
```

## Arguments

data	FELLA.DATA object
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"
squared	Logical, whether to return rowSums (F) or squaredRowSums (T) $% \left( T\right) =\left( T\right) \left( T\right)$

## Value

Named vector with rowSums/squaredRowSums (internal usage)

## Examples

```
## This function is internal
attach(environment(FELLA:::getSums))
data(FELLA.sample)
rowsums <- getSums(FELLA.sample, "diffusion", squared = FALSE)
hist(rowsums)</pre>
```

getValid

Get the slot "valid"

## Description

Extractor function for the slot "valid"

## Usage

```
getValid(object, method)
```

## Arguments

object	FELLA.USER object
method	Character, exactly one of: "hypergeom", "diffusion", "pagerank"

## Value

Slot "valid" (internal usage)

## Examples

## This function is internal

data(FELLA.sample)
data(input.sample)

obj <- enrich( compounds = input.sample, method = "diffusion", data = FELLA.sample)

## If the analysis is valid
FELLA:::getValid(obj, "diffusion")

## Otherwise
FELLA:::getValid(new("FELLA.USER"), "diffusion")
FELLA:::getValid(obj, "pagerank")

infere.con2ec Infer connections to EC

## Description

Function infere.con2ec infers network connections to KEGG EC families by passing through genes. This assumes that the category being mapped to enzymes is above them.

## Usage

infere.con2ec(ids, ent, ent2gene, gene2enzyme)

## Arguments

ids	Character vector of identifiers to map. For example, all the KEGG pathways
ent	Character, entity that we are mapping (one of "pathway" and one of "module")
ent2gene	Named character vector, names are the entity ent and values are genes
gene2enyzme	Named character vector, names are genes and values are EC enzyme families category Character, one of:

#### Value

Two-column data frame. Column "from" contains the KEGG enzyme families whereas "to" contains the entity ent.

## input.sample

#### Examples

```
ids <- "hsa00010"
ent <- "pathway"
ent2gene <- c("hsa00010" = "hsa:10", "hsa00010" = "hsa:120")
gene2enzyme <- c("hsa:10" = "1.1.1.1", "hsa:120" = "1.2.3.4")
FELLA:::infere.con2ec(ids, ent, ent2gene, gene2enzyme)</pre>
```

input.sample

```
A randomly generated list of affected metabolites
```

#### Description

This character vector object has been generated using the sample data in the object FELLA. sample. The KEGG compounds have been chosen with preference for the hsa00640 pathway, so that the enrichment results choose pathway hsa00640 over hsa00010.

## Usage

```
data(input.sample)
```

#### Format

An object of class character of length 30.

#### Value

A character vector containing 30 KEGG IDs

#### Source

Generated from a mid-2017 KEGG release (http://www.genome.jp/kegg/)

## Examples

data(input.sample)

is.FELLA.DATA

## Description

Is x a FELLA. DATA object?

#### Usage

is.FELLA.DATA(x = NULL)

## Arguments

х

Object to check

## Value

Logical value stating if x is a FELLA. DATA object

## Examples

data(FELLA.sample)
is.FELLA.DATA(FELLA.sample)
is.FELLA.DATA(42)

is.FELLA.USER Check FELLA.USER class

## Description

Is x a FELLA. USER object?

#### Usage

is.FELLA.USER(x = NULL)

# Arguments ×

Object to check

#### Value

Logical value stating if x is a FELLA. USER object

## largestcc

## Examples

```
is.FELLA.USER(new("FELLA.USER"))
is.FELLA.USER(42)
data(FELLA.sample)
data(input.sample)
obj <- enrich(
  compounds = input.sample,
  method = "diffusion",
  data = FELLA.sample)
is.FELLA.USER(obj)</pre>
```

largestcc

Extract largest CC

## Description

Function largestcc extracts the largest connected component of an igraph object

## Usage

largestcc(graph)

## Arguments

graph Igraph object

#### Value

Connected igraph object

## Examples

```
library(igraph)
g <- barabasi.game(10) + graph.empty(10)
FELLA:::largestcc(g)</pre>
```

launchApp

#### Description

launchApp deploys a shiny application to perform the metabolomics data enrichment. Although this app does not provide all the options available in FELLA, it is easily accessible for the lay user.

## Usage

launchApp(...)

#### Arguments

... Parameters passed to runApp

## Details

The graphical interface allows to: (1) upload the data and check if the KEGG ids have successfully mapped, (2) select database, set analysis and graphical parameters, (3) interactively browse the resulting sub-network as a graph or as a table, and (4) export such results as a table or a network. At least one database is needed before deploying the app. See ?buildDataFromGraph for further details.

#### Value

invisible(), but as a side effect the app will be launched

#### Examples

```
## Not run:
r <- try(launchApp())
```

## End(Not run)

listApprox

#### List of approximations

#### Description

Available approximations for the analysis

#### Usage

listApprox()

# listCategories

## Value

Character vector

## Examples

listApprox()

listCategories List of node categories

## Description

Node categories used in the internal representations

## Usage

listCategories()

#### Value

Character vector

## Examples

listCategories()

listInternalDatabases List internal databases

## Description

This function lists the directories in the local database path

## Usage

```
listInternalDatabases(full.names = FALSE)
```

## Arguments

full.names Logical, should full paths be returned?

#### Value

Vector with database directories

## Examples

listInternalDatabases()

listMethods List of methods

## Description

Methods available for the analysis

## Usage

listMethods()

## Value

Character vector

## Examples

listMethods()

mytriangle

Add triangular shape to igraph plot

## Description

This function enables the usage of triangles as shape in the function plot.igraph.

## Usage

mytriangle(coords, v = NULL, params)

# Arguments

coords, v, params

clipping arguments, see shapes

## Value

Plot symbols

40

## plotBipartite

#### Examples

```
## This function is internal
library(igraph)
add.vertex.shape(
  "triangle", clip = shapes("circle")$clip,
plot = FELLA:::mytriangle)
g <- barabasi.game(10)
plot(
g, vertex.shape = "triangle",
vertex.color = rainbow(vcount(g)),
vertex.size = seq(10, 20, length = vcount(g)))
```

plotBipartite Internal function to plot a bipartite graph

#### Description

This function plots a bipartite graph, tailored for the hypergeometric over representation analysis. As the nodes can only be compounds and pathways, the layout is simple and bipartite.

#### Usage

```
plotBipartite(graph = NULL, layout = FALSE, ...)
```

#### Arguments

graph	Graph result that must come from the hypergeometric test analysis
layout	Logical, should the plot be returned as a layout?
	Additional parameters passed to plot.igraph

#### Value

If layout = F then the value returned is invisible(). Otherwise, the layout is returned, also in an invisible fashion.

## Examples

```
## This function is internal
data(FELLA.sample)
data(input.sample)
## Enrich input
obj <- enrich(
compounds = input.sample,
data = FELLA.sample,
method = "hypergeom")</pre>
```

## plotLegend

```
## Generate the bipartite graph (only in the hypergeometric test)
g <- generateResultsGraph(
method = "hypergeom",
threshold = 1,
object = obj,
data = FELLA.sample)
## Plot it
FELLA:::plotBipartite(g)</pre>
```

plotLegend

## Internal function to add a legend to a graph plot

## Description

This function adds a legend to a solution plot. It can include the CC similarity.

## Usage

```
plotLegend(GO.annot = FALSE, cex = 0.75)
```

## Arguments

GO.annot	Logical, should GO annotations be included?
cex	Numeric value, cex parameter for the function legend

#### Value

This function is only used for its effect, so it returns invisible()

## Examples

```
## This function is internal
```

```
library(igraph)
g <- barabasi.game(20)
plot(g)
FELLA:::plotLegend()
plot(g)
FELLA:::plotLegend(G0.annot = TRUE)</pre>
```

42

sanitise

## Description

Sanitise KEGG identifiers

#### Usage

sanitise(x, category, organism)

## Arguments

х	Character vector, IDs to sanitise
category	Character, one of: "pathway", "module", "enzyme", "ncbi", "reaction", "compound"

## Value

Character vector, sanitised x

## Examples

```
FELLA:::sanitise(c("path:hsa00010", "path:hsa00020"), "pathway", "hsa")
```

U.diffusion-class	An internal S4 class for the user data of the diffusion enrichment anal-
	ysis

## Description

An internal S4 class for the user data of the diffusion enrichment analysis

#### Slots

valid Logical value; is the analysis valid?

pscores Named numeric vector with p-scores

- approx Character; which approximation was used? Can be "simulation" for Monte Carlo; "normality", "gamma" or "t" for parametric approaches
- niter Numeric value, number of iterations for the simulated approach

U.hypergeom-class

#### Description

An internal S4 class for the user data of the hypergeometric over representation analysis

#### Slots

valid Logical value; is the analysis valid?

pvalues Named numeric vector with p-values

- pathhits Numeric named vector with the quantities "sample\_success" for the hypergeometric distribution (#affected in path)
- pathbackground Numeric named vector with the quantities "total\_success" for the hypergeometric distribution (total in path)
- nbackground Numeric value, number of compoudns in the background. Equivalently, number of rows for the hypergeometric binary matrix
- ninput Numeric value, number of affected compounds matched to the rownames
- U.pagerank-class An internal S4 class for the user data of the PageRank enrichment analysis

#### Description

An internal S4 class for the user data of the PageRank enrichment analysis

#### Slots

valid Logical value; is the analysis valid?

pscores Named numeric vector with p-scores

- approx Character; which approximation was used? Can be "simulation" for Monte Carlo; "normality", "gamma" or "t" for parametric approaches
- niter Numeric value, number of iterations for the simulated approach

U.userinput-class An internal S4 class for the user input data

## Description

An internal S4 class for the user input data

#### Slots

metabolites Character vector containing the affected compounds

- metabolitesbackground Character vector containing the compounds for the personalised background. Optionally, can be NULL for default background
- excluded Character vector containing the compounds that have been excluded because they cannot be mapped to KEGG graph compounds

# Index

```
* datasets
    FELLA.sample, 23
    input.sample, 35
* internal
    .params, 3
    checkArguments, 5
    getPvaluesSize, 31
    infere.con2ec, 34
    largestcc, 37
    mytriangle, 40
    plotBipartite, 41
    sanitise, 43
.params, 3
addGOToGraph (export-funs), 16
buildDataFromGraph, 19
buildDataFromGraph(data-funs), 8
buildGraphFromKEGGREST(data-funs), 8
checkArguments, 5
D.diffusion-class, 6
D.hypergeom-class, 7
D.keggdata-class, 7
D.pagerank-class, 7
data-funs, 8
defineCompounds (enrich-funs), 11
enrich (enrich-funs), 11
enrich-funs, 11
export-funs, 16
exportResults (export-funs), 16
FELLA, 9, 12, 21, 38
FELLA-package (FELLA), 21
FELLA.DATA, 8–15, 18, 19, 23, 36
FELLA.DATA (FELLA.DATA-class), 22
FELLA.DATA-class, 22
FELLA.sample, 23
FELLA.USER, 9, 11-15, 17, 18, 24, 36
```

FELLA.USER (FELLA.USER-class), 24 FELLA.USER-class, 24 generateEnzymesTable (export-funs), 16 generateResultsGraph (export-funs), 16 generateResultsTable (export-funs), 16 getBackground, 25 getBM, 4, 18 getCom, 26 getExcluded, 27 getGraph, 27 getInfo, 28 getInput, 29 getMatrix, 29 getName, 30 getPscores, 31 getPvaluesSize, 31 getStatus, 32 getSums, 33 getValid, 33 godata, 4, 18 infere.con2ec.34 input.sample, 35 is.FELLA.DATA, 36 is.FELLA.USER, 36 largestcc, 37 launchApp, 22, 38, 38 legend, 42 listApprox, 38 listCategories, 39 listDatasets, 4, 18 listInternalDatabases, 39 listMethods, 40 loadKEGGdata(data-funs), 8 mytriangle, 40 p.adjust, 4, 12 page.rank, 4, 5, 8, 12, 13

## INDEX

```
pgamma, 14
plot,FELLA.USER,missing-method
(FELLA.USER-class), 24
plot.igraph, 19, 40, 41
plotBipartite, 41
plotGraph (export-funs), 16
plotLegend, 42
pnorm, 14
pt, 14
```

```
runApp, 38
runDiffusion(enrich-funs), 11
runHypergeom(enrich-funs), 11
runPagerank(enrich-funs), 11
```

```
sanitise, 43
shapes, 40
show,FELLA.DATA-method
    (FELLA.DATA-class), 22
show,FELLA.USER-method
    (FELLA.USER-class), 24
```

tkplot, 19

```
U.diffusion-class, 43
U.hypergeom-class, 44
U.pagerank-class, 44
U.userinput-class, 45
```

write\_graph, *18*, *19*